Diphtheria

Signs and Symptoms

Diphtheria is a serious infection caused by toxigenic strains of bacteria called *Corynebacterium diphtheriae*. It can infect any mucus membrane; symptoms of diphtheria depend on the body part that is affected. Early symptoms of toxigenic respiratory infection include malaise, sore throat, anorexia, and low-grade fever (less than 101°F). A blush-white “pseudomembrane” forms and extends varying in size from covering a small patch on the tonsils to covering most of the soft palate.

Non-toxigenic *C. diphtheriae* infections can still cause disease, including wound infections, bacteremia, and sometimes even a less serious respiratory illness; however, non-toxigenic infection is not the same as diphtheria disease.

*Complications: pneumonia, myocarditis, endocarditis, neuritis, airway obstruction, septic arthritis, osteomyelitis, and death. The severity of symptoms correlates with the location and extent of the local disease*

Incubation

For toxigenic diphtheria: 1 to 10 days; average 2 to 5 days.

Case Classification

**Clinical definition:** Upper respiratory tract illness with an adherent membrane of the nose, pharynx, tonsils, or larynx OR Infection of a non-respiratory anatomical site (e.g., skin, conjunctiva, ear, genital mucosa)

**Confirmed case:** meets clinical definition AND
- laboratory confirmed OR
- epi-linked to a lab-confirmed case of diphtheria

**Suspected case:** meets clinical definition AND absence of laboratory confirmation AND lack of epi-linkage to a lab-confirmed case of diphtheria OR histopathologic diagnosis

Differential diagnosis

Pharyngitis, Infectious Mononucleosis, Oral Syphilis, Epiglottitis, Oral Candidiasis, Angioedema

Treatment

Administration of Diphtheria Antitoxin (DAT), available at CDC quarantine stations and antimicrobial therapy (typically erythromycin or penicillin) to kill the bacteria

**Culture:** Clinical specimens for isolation and identification of *C. diphtheriae*. PHL performs this test.

**Identification:** *C. diphtheriae* isolates can be confirmed using traditional biochemical methods at PHL. *All C. diphtheriae isolates, regardless of association with disease, are required to be sent to PHL. They will be submitted to CDC where testing will be performed to determine if the isolate is producing diphtheria toxin. (See Section 4.A.)*

Laboratory investigation

- Assess the likelihood of (toxigenic) diphtheria: confirm compatible clinical symptoms, verify vaccination and travel history, and assess exposure risk such as contact with a person with diphtheria or recent visit(s) to a healthcare facility.
- Recommend immediate isolation of case (droplet precautions for suspected respiratory diphtheria and contact precautions for suspected cutaneous diphtheria) until diphtheria is ruled out, or 2 cultures taken at least 24 h apart and 24 h after antimicrobial therapy cessation are negative.
- Consult with DOH CD Epi regarding the need for DAT, and specimen collection for confirmatory/toxigenicity testing. Keep in mind that toxigenicity testing at CDC may take 2 weeks or longer from the first positive result.
- Recommend appropriate infection control precautions to prevent additional exposures in healthcare facilities, schools, and other public settings.
- Identify close contacts and determine their immune status. Close contacts of a confirmed case should have cultures taken from nose and throat regardless immunization status or the presence of symptoms, and they should receive antimicrobial therapy.
- Recommend vaccination for all cases during the convalescence period (disease does not always confer immunity), and for any close contact whose last dose was received more than 5 years ago.
1. DISEASE REPORTING

A. Purposes of Reporting and Surveillance

1. To assist in the identification of cases.
2. To assure early and appropriate treatment with diphtheria antitoxin and antibiotics along with isolation to prevent transmission if needed.
3. To identify and evaluate contacts and recommend appropriate antibiotic prophylaxis and/or immunization to prevent further spread of the disease.
4. To alert public health authorities and health care providers to the presence of diphtheria in the community and the potential for additional infections, a particular concern given the large number of susceptible adults.

B. Legal Reporting Requirements

1. Health care providers and Health Care Facilities: immediately notifiable to local health jurisdiction.
2. Laboratories: *Corynebacterium diphtheriae* immediately notifiable to local health jurisdiction, submission required – isolate within 2 business days; submission on request – specimen associated with positive result, within 2 business days.
3. Local health jurisdictions: immediately notifiable to the Washington State Department of Health (DOH) Office of Communicable Disease Epidemiology (CDE) at 877-539-4344 or 206-415-5500.

C. Local Health Jurisdiction Investigation Responsibilities

1. Begin case investigation immediately. If the health care provider requests antitoxin contact CDE to facilitate release of the biologic.
2. Facilitate the transport of specimens to assist with the diagnosis.
3. Recommend measures to prevent further spread from the case.
4. Identify and evaluate contacts; educate and recommend measures to prevent further spread from contacts.
5. Report all confirmed and suspect cases (see Section 3C) to CDE. Complete the diphtheria case report form [http://www.doh.wa.gov/Portals/1/Documents/5100/210-056-ReportForm-Diphtheria.pdf](http://www.doh.wa.gov/Portals/1/Documents/5100/210-056-ReportForm-Diphtheria.pdf) and enter the data into the Washington Disease Reporting System (WDRS).

2. THE DISEASE AND ITS EPIDEMIOLOGY

A. Etiologic Agent

Diphtheria is an acute bacterial disease caused by toxin-producing strains of *Corynebacterium diphtheriae*. The toxin causes local tissue destruction and membrane formation. It primarily affects the tonsils, pharynx, nose and larynx. Other mucous membranes, skin, and rarely the vagina or conjunctivae can also be involved.
Susceptible persons may acquire toxigenic diphtheria bacilli in the nasopharynx. The organism produces a toxin that inhibits cellular protein synthesis and is responsible for local tissue destruction and formation of the pseudomembrane that is characteristic of this disease. The toxin produced at the site of the membrane is absorbed into the bloodstream and then distributed to the tissues of the body. The toxin is responsible for major complications such as myocarditis, polyneuropathies, and nephritis, and thrombocytopenia.

Complications of diphtheria include pneumonia, myocarditis, neuritis, airway obstruction, septic arthritis, osteomyelitis, and death. The case-fatality rate for classic diphtheria is approximately 10%.

For clinical purposes, diphtheria can be classified according to the site of the infection:

1. **Pharyngeal and tonsillar diphtheria (faucial)**

   Pharyngeal and tonsillar diphtheria is the most common type of toxigenic infection (about 70% of cases). It initially presents with an insidious onset of malaise, sore throat, anorexia, and low-grade fever (usually under 102 F). At onset of symptoms, the pharynx is erythematous, but no membrane is present. About a day after onset, small patches of exudate appear in the pharynx. Despite the low-grade fever, patients usually appear quite ill and have tachycardia (“toxic appearance”). Within 2 or 3 days of onset, the patches of exudate spread and become confluent and may form a bluish-white pseudomembrane that can extend to the entire pharynx, including the nasopharynx, tonsillar areas, soft palate, and uvula. This membrane becomes grayish, thick, firmly adherent, and may have patches of green or black necrosis. Attempts to remove the pseudomembrane can cause bleeding.

   Enlargement and tenderness of the anterior cervical lymph nodes is common. With severe disease, patients can develop edema of the soft tissues in the anterior neck, giving a characteristic “bullneck” appearance, which can cause respiratory stridor and is associated with a higher morbidity and mortality.

   The severity of symptoms correlates with the location and extent of the membrane. In untreated patients, the membrane begins to soften about a week after onset and gradually detaches, usually in pieces. As the membrane detaches, acute systemic symptoms disappear. However, at any point during the course of the illness, if a significant amount of toxin is absorbed into the blood stream, patients may develop pallor and a rapid pulse which can progress to coma and even death.

2. **Laryngeal diphtheria**

   Laryngeal diphtheria occurs in about 25% of cases and is more likely to occur in children younger than 4 years. When the infection involves the larynx, it may occur either as an extension of the pharyngeal form, or as laryngeal involvement alone. Patients can present with fever, hoarseness, dyspnea, respiratory stridor, and a barking cough. In this form, also, the pseudomembrane can cause potentially fatal airway obstruction and a greater degree of toxin absorption if the membrane is extensive.
3. **Anterior nasal diphtheria**

Anterior nasal diphtheria usually presents with mucopurulent discharge from the nose, which may be bloody, and is often associated with a white pseudomembrane on the septum. External nares and upper lip may also be involved. Anterior nasal diphtheria as the only manifestation is uncommon (about 2% of cases).

The differential diagnosis of respiratory diphtheria (i.e. pharyngeal, laryngeal, anterior nasal) includes infection with other pathogens that can cause similar symptoms including: other Corynebacterium species, Arcanobacterium haemolyticum, as well as Streptococcus spp., Epstein-Barr virus and cytomegalovirus (both of which cause infectious mononucleosis syndrome). Candida albicans, syphilis, bacterial anaerobes (such as the organisms associated with Vincent’s angina). Some viruses may cause a membrane of the throat and tonsils as well.

4. **Cutaneous (skin) diphtheria**

Cutaneous diphtheria, caused by toxin-producing *C. diphtheriae* is usually mild, typically consisting of indistinct sores or shallow ulcers. Toxigenic strains appear to result in less systemic complications compared to other forms of diphtheria.

Cutaneous diphtheria may act as a reservoir for transmission and result in respiratory or cutaneous infections in other susceptible hosts. Thorough cleansing of the lesion with soap and water and appropriate antimicrobial therapy are recommended.

Other possible sites of infection include the conjunctiva, vulvovaginal area, and external auditory canal. Severe disease is more likely to occur in people who are unimmunized or under immunized.

5. **Carrier**

In some people, infection with diphtheria-causing bacteria causes only a mild illness, or no obvious signs and symptoms at all. Infected people who are asymptomatic are known as carriers because they can spread the infection without being sick themselves.

6. **Non-toxigenic *C. diphtheriae* infection**

Non-toxin-producing strains of *C. diphtheriae* have been increasingly isolated from wounds, blood, and respiratory sites. These non-toxigenic strains can cause an infection which is generally less severe, potentially causing a mild sore throat and, rarely, membranous pharyngitis in the respiratory tract. Invasive disease, including bacteremia and endocarditis, has been reported.

*C. diphtheriae* isolated from cutaneous cases in the United States typically has been nontoxigenic. Any chronic skin lesion may harbor *C. diphtheriae* along with other organisms. Some cutaneous infections have led to major complications including amputations and death.

**B. Reservoir**

Humans
C. Modes of Transmission

Transmission is most often person-to-person spread from the respiratory tract. Rarely, transmission may occur from skin lesions or articles soiled with discharges from lesions of infected persons (fomites).

D. Incubation Period

The incubation period is usually 2–5 days (range, 1–10 days).

E. Period of Communicability

Once infected, untreated persons can shed bacteria from the respiratory tract or from skin lesions for 2–6 weeks. Once an effective antibiotic has been initiated, persons are communicable for up to 4 days. Isolation should be maintained until two cultures have shown an absence of the organism (See Section 6.A.3). A carrier may shed organisms for 6 months or more, but effective antibiotic therapy eliminates shedding.

F. Treatment

The treatment for toxigenic diphtheria is prompt administration of diphtheria antitoxin (DAT). If toxigenic diphtheria is strongly suspected on the basis of clinical and epidemiologic findings, specimens for bacteriologic testing should be collected, then antitoxin given as soon as possible, without waiting for test results. Toxigenicity test results can take 2 weeks or longer to be available, waiting for results before treatment is not possible.

CDC maintains a supply of DAT at quarantine stations around the country, including the one located at SeaTac airport. DAT is currently available for treatment of respiratory diphtheria under an FDA-approved Investigational New Drug (IND) protocol. Since the antitoxin is of equine origin, a test to rule out hypersensitivity should be performed before administration. Antitoxin may only be administered in an inpatient environment.

**Healthcare providers of a patient with suspected diphtheria should contact their local health jurisdiction immediately.** The local health jurisdiction, in collaboration with DOH, will arrange a consultation with CDC, and subsequent transport of antitoxin if needed. For additional information regarding DAT, see:


The recommended antibiotics for diphtheria are erythromycin or penicillin G. Antibiotics must be initiated as soon as possible for the eradication of the organism, help limit the toxin released into the system, quicken the recovery phase in the patient, and prevent the spread of the infection to close contacts. In the case of antibiotic resistance, linezolid or vancomycin can be used.

Treatment of cutaneous diphtheria with antibiotics is usually sufficient, antitoxin is typically not needed.

Appropriate isolation precautions should remain in place until diphtheria is ruled out or until off antimicrobial treatment and culture negative (See Section 6.A.3).

If not immunized, carriers should receive active immunization and measures should be taken to ensure completion of the immunization schedule. If a carrier has been immunized previously but has not received a booster of diphtheria toxoid within 5 years, a booster dose of age-appropriate vaccine should be administered. Carriers should receive
oral erythromycin or a single intramuscular dose of penicillin G benzathine. Two follow-up cultures should be performed after completing antimicrobial treatment to detect persistence of carriage, which occurs following erythromycin treatment in some cases. The first culture should be performed 24 hours after completing treatment. If results of cultures are positive, an additional 10-day course of oral erythromycin should be administered, and follow-up cultures should be performed again.

**G. Immunity**

A protective level of antitoxin (defined as greater than 0.1 IU of antitoxin/mL) is reached in more than 95% of vaccine recipients. Diphtheria toxoid-containing vaccine has been estimated to have an efficacy of 97%. After a primary series of 3 properly spaced doses of diphtheria toxoid-containing vaccines in infants and a booster dose at age 15 through 18 months or 3 properly spaced doses in adults.

*Use of Tetanus Toxoid, Reduced Diphtheria Toxoid, and Acellular Pertussis Vaccines: Updated Recommendations of the Advisory Committee on Immunization Practices—United States, 2019 | MMWR (cdc.gov)*

Diphtheria disease might not confer immunity. Unvaccinated or incompletely vaccinated persons recovering from diphtheria should begin or complete active immunization with diphtheria toxoid during convalescence.

### 3. CASE AND CONTACT DEFINITIONS

**A. Clinical description**

Classic diphtheria is an upper-respiratory tract illness characterized by sore throat, low-grade fever, and an adherent pseudomembrane on the tonsil(s), pharynx, larynx, and/or nose. However, disease can involve almost any mucous membrane. For clinical purposes it is convenient to classify diphtheria depending on the site of disease:

- anterior nasal diphtheria
- pharyngeal and tonsillar diphtheria
- laryngeal diphtheria
- cutaneous (skin) diphtheria

**B. Laboratory criteria for diagnosis**

Confirmatory laboratory evidence:

- Isolation of *Corynebacterium diphtheriae* from a clinical specimen, AND
- Confirmation of toxin-production by Elek test or by another validated test capable of confirming toxin-production

Supportive laboratory evidence:

- Histopathologic diagnosis of diphtheria

**C. Case definition (2019)**

1. Suspect: In the absence of a more likely diagnosis, an upper respiratory tract illness with each of the following:

- an adherent membrane of the nose, pharynx, tonsils, or larynx, AND
- absence of laboratory confirmation, AND
• lack of epidemiologic linkage to a laboratory-confirmed case of diphtheria;

OR

• Histopathologic diagnosis

2. Confirmed:

• An upper respiratory tract illness with an adherent membrane of the nose, pharynx, tonsils, or larynx, and any of the following:
  - isolation of toxin-producing C. diphtheriae from the nose or throat, OR
  - epidemiologic linkage to a laboratory-confirmed case of diphtheria;

OR

• An infection at a non-respiratory anatomical site (e.g., skin, wound, conjunctiva, ear, genital mucosa) with isolation of toxin-producing C. diphtheriae from that site.

An epidemiologically linked case requires direct contact with a laboratory-confirmed case of diphtheria.

D. Comment

All C. diphtheriae isolates, regardless of association with disease, should be submitted to the Washington State Public Health Laboratories (PHL) for submission to the Diphtheria Laboratory at CDC.

Rarely, respiratory diphtheria may result from infection with other Corynebacterium species (C. ulcerans or C. pseudotuberculosis). These bacteria, if isolated from someone with a diphtheria-like illness, should also be forwarded to PHL as it is possible for them to produce diphtheria toxin, and they can be tested for toxigenicity using the Elek test at CDC.

4. DIAGNOSIS AND LABORATORY SERVICES

A. Diagnosis

The initial diagnosis of diphtheria should be based on the clinical presentation and epidemiologic risk factors because it is imperative to begin presumptive therapy quickly.

Culture and toxigenicity testing: Diphtheria is confirmed by isolation of Corynebacterium diphtheriae followed by toxigenicity testing. When diphtheria is suspected, the clinical laboratory receiving specimens for testing should be advised so that culture medium that provides a selective advantage for the growth of C. diphtheria can be used, if available.

When C. diphtheriae is identified, an isolate must be tested for expression of diphtheria toxin by the Elek test method at CDC, because the presence of the tox gene does not necessarily indicate that toxin is being produced. The Elek test is a functional test which shows whether the bacteria is actively producing toxin.

Serologic testing: Serum antibody levels can assist with assessing the likelihood of the diagnosis whenever diphtheria is suspected. Specimens must be collected prior to the administration of DAT. When antibiotics were administered prior to collection of
specimens for culture, health care providers should be strongly encouraged to obtain a serum specimen.

- Serum antibody levels less than 0.01 IU/ml – likely susceptible to diphtheria
- Serum antibody levels between 0.01–0.09 IU/ml – indicates basic immunity.
- Serum antibody levels ≥0.10 IU/ml – considered fully protective.

Testing for serum antibody levels is available at commercial laboratories.

B. Tests Available at Washington State Public Health Laboratories (PHL)

The primary role of the PHL Special Bacteriology lab is to confirm the identification of *C. diphtheriae* isolates already obtained at a community laboratory, PHL can also perform the initial culture to see if *C. diphtheriae* is present in a clinical specimen.

All requests for diphtheria testing to be done at PHL or forwarded to CDC for testing must have approval from an CDE epidemiologist prior to specimen submission.

Note that PHL requires all clinical specimens have two patient identifiers, a name and a second identifier (e.g., date of birth) both on the specimen label and on the submission form. Due to laboratory accreditation standards, specimens will be rejected for testing if not properly identified. Include specimen source and collection date.

C. Specimen Collection

Information on specimen collection and submission instructions for *C. diphtheriae* culture and identification can be found at [PHL Microbiology testing menu](https://www.medialab.com/dv/dl.aspx?d=1887088&dh=52b1a&u=69790&uh=0e2a1).

Culture specimens: Obtain a culture from the nose and throat of all persons with suspected diphtheria (nasopharyngeal swab preferred). If possible, swabs should also be taken from beneath the adherent pseudomembrane, and a portion of the pseudomembrane collected in a sterile container. For suspected cutaneous diphtheria, a swab of the infected area is the preferred specimen. Specimens for culture should be obtained as soon as diphtheria (involving any site) is suspected, even if treatment with antibiotics has already begun.

Clinical specimens should reach the PHL as quickly as possible after collection.

If no diphtheria isolate can be obtained from a patient receiving DAT, a clinical specimen should be sent to PHL for direct testing at CDC for the presence of the tox gene using PCR.

Collection of clinical specimens for isolation of *C. diphtheriae* from close contacts (potential carriers) of a highly suspected diphtheria case can also aid in the presumptive diagnosis.

Presumptive *C. diphtheriae* isolates and clinical specimens are required to be submitted with a completed DOH microbiology form available at: [https://www.medialab.com/dv/dl.aspx?d=1887088&dh=52b1a&u=69790&uh=0e2a1](https://www.medialab.com/dv/dl.aspx?d=1887088&dh=52b1a&u=69790&uh=0e2a1)

For additional information regarding laboratory testing for diphtheria, see: [https://www.cdc.gov/vaccines/pubs/surv-manual/chpt01-dip.html](https://www.cdc.gov/vaccines/pubs/surv-manual/chpt01-dip.html)
5. ROUTINE CASE INVESTIGATION

A. Evaluate the Diagnosis

Review the clinical presentation, risk factors for exposure, and immunization status to determine the likelihood of the diagnosis.

If diphtheria is highly suspected, do the following:

- Assure that the patient is in an isolation room with standard plus droplet precautions.
- Immediately consult with CDE regarding the need for testing and treatment with diphtheria antitoxin. CDE will facilitate CDC consultation as needed.
- Request collection of specimens for confirmation of the diagnosis at PHL. Collect serum to be held for serologic testing, if needed.
- Recommend the initiation of antibiotic treatment. Treatment should not be delayed pending laboratory confirmation when the diagnosis of diphtheria is strongly suspected. Erythromycin or penicillin G are the recommended antibiotics.
- Facilitate the transportation of specimens to PHL.

If the suspicion of diphtheria is low, specimens can be sent to a commercial laboratory, laboratory staff should be alerted that diphtheria is included in the differential diagnosis.

B. Identify Source of Infection

Ask the patient about potential sources of infection in the 10 days prior to onset including:

- Travel out of the country, especially to an area where diphtheria is still endemic.
- Contact with persons from a country where diphtheria is still endemic; and
- Working, visiting, or volunteering in a healthcare setting.

*Please note: Using nose and throat cultures to search for diphtheria carriers, other than among close contacts, is not ordinarily useful or indicated.*

C. Identify Close Contacts

Identify all close contacts, particularly household members and others who were directly exposed to respiratory secretions of the case and determine their immunization status. See section 6.B. for managing contacts.

D. Environmental evaluation

None

6. CONTROLLING FURTHER SPREAD

A. Infection Control Recommendations/Case Management

1. Persons with confirmed pharyngeal diphtheria should be cared for using standard plus droplet precautions until they have completed antimicrobial therapy and two cultures
taken at least 24 hours apart, and at least 24 hours after cessation of antimicrobial therapy, fail to show diphtheria organisms.

2. Persons with cutaneous diphtheria should be cared for using standard plus contact precautions until they have completed antimicrobial therapy and two cultures taken at least 24 hours apart, and at least 24 hours after cessation of antimicrobial therapy, fail to show diphtheria organisms. Wounds should remain covered until drainage stops or can be contained by a dressing.

3. Persons with confirmed diphtheria should avoid close contact with others until two cultures taken 24 hours apart, and at least 24 hours after cessation of antimicrobial therapy, fail to show diphtheria organisms.

4. Persons with diphtheria should be vaccinated with diphtheria toxoid during convalescence because clinical disease does not necessarily confer immunity.

B. Contact Management

An investigation for all suspected respiratory and non-respiratory diphtheria cases. Cases of cutaneous or respiratory diphtheria caused by infections with nontoxigenic strains of C diphtheriae are not nationally notifiable and do not require routine investigation or prophylaxis of contacts; however, some case investigation is recommended to determine the likelihood of the infection being toxigenic (e.g., case had international travel or respiratory symptoms like diphtheria). Close contacts of a person suspected to have diphtheria should be identified, and the following are recommended:

1. Contact tracing usually can be limited to household members and people with direct, habitual close contact or health care personnel exposed to nasopharyngeal secretions, people sharing kitchen facilities, or people caring for infected children.

2. Close contacts with symptoms compatible with diphtheria should be referred to a health care provider for evaluation immediately.

3. Surveillance for evidence of disease in all is necessary for 7-10 days from last exposure to an untreated patient.

4. Culture all close contacts for C. diphtheriae. Specimens should be obtained from nares and throat or any mucosal or cutaneous lesion.

5. Close contacts, regardless of their immunization status, should receive erythromycin or penicillin as prophylaxis. Follow-up cultures of pharyngeal specimens should be performed after completion of therapy for contacts proven to be carriers. If cultures are positive, an additional 10-day course of erythromycin should be administered, and follow-up cultures of pharyngeal specimens again should be performed.

6. For compliance reasons, if the health department cannot maintain surveillance of close contacts, the close contacts should receive a dose of intramuscular benzathine penicillin. If not fully immunized or if immunization status is not known, they should be immunized with diphtheria-containing vaccine as appropriate for age.

7. Asymptomatic, previously immunized close contacts should receive a booster dose of an age-appropriate diphtheria toxoid-containing vaccine if they have not received a booster dose of a diphtheria toxoid-containing vaccine within 5 years.

8. Asymptomatic close contacts who have had fewer than 3 doses of a diphtheria toxoid-
containing vaccine, children younger than 7 years in need of their fourth dose of DTaP (or DT), or people whose immunization status is not known should be immunized with an age-appropriate diphtheria toxoid-containing vaccine.

9. Use of equine diphtheria antitoxin in unimmunized close contacts is not recommended, because there is no evidence that antitoxin provides additional benefit.

10. Close contacts should watch for symptoms of diphtheria during the period from the day after the first possible exposure through 10 days after the last known exposure. Daily symptom check by public health should be considered for contacts that were unimmunized when exposed shown to be nontoxigenic, the health department can discontinue investigation of contacts.

For additional information regarding case investigations, see the CDC VPD Surveillance Manual available at: http://www.cdc.gov/vaccines/pubs/surv-manual/chpt01-dip.html

C. Environmental measures

None

7. MANAGING SPECIAL SITUATIONS

Special situations will be handled on a case-by-case basis. Please consult with Office of Communicable Disease Epidemiology.

8. ROUTINE PREVENTION

A. Immunization Recommendations

Diphtheria, tetanus, and acellular pertussis vaccination is recommended across the lifespan. Children younger than 7 years of age receive DTaP or DT, while older children and adults receive Tdap and Td.

- Infants and children receive 5 doses of DTaP. Give one dose at each of these ages: 2 months, 4 months, 6 months, 15 through 18 months, and 4 through 6 years. Use DT for infants and children who should not receive acellular pertussis-containing vaccines.
- Adolescents receive a single dose of Tdap, preferably at 11 to 12 years of age.
- A single dose of Tdap is given during every pregnancy, preferably during the early part of gestational weeks 27 through 36.

Routine booster doses of Td or Tdap vaccine should be given at 10-year intervals.

The full routine vaccination schedule and catch-up recommendations are available at: https://www.cdc.gov/vaccines/schedules/hcp/imz/child-adolescent.html

For additional information regarding use of the diphtheria vaccines, adverse reactions and contraindications see the most recent CDC Pink Book available at: http://www.cdc.gov/vaccines/pubs/pinkbook/dip.html

B. Prevention Recommendations

Immunization is the best way to prevent diphtheria.
ACKNOWLEDGEMENTS
This document is a revision of the Washington State Guidelines for Notifiable Condition Reporting and Surveillance published in 2002 which were originally based on the Control of Communicable Diseases Manual (CCDM), 17th Edition; James Chin, Ed. APHA 2000. We would like to acknowledge the Oregon Department of Human Services for developing the format and select content of this document.

UPDATES

January 2011: The Legal Reporting Requirements section has been revised to reflect the 2011 Notifiable Conditions Rule revision. Updated to include the 2010 CSTE case classification changes.

March 2016: Clinical presentation and Epidemiology sections were reviewed and updated according to the most recent medical literature available. Specimen collection section was updated to reflect current testing available at PHL and CDC. Immunization section was updated for consistency with current ACIP recommendations.

December 2022: For 2023 WAC revision combined provider and facility reporting requirement, updated laboratory submission (Section 1B)

September 2023: CSTE case definition was updated in 2019, updating guideline to reflect this update. Expanded information provided for cutaneous diptheria. Updated recommendations for close contacts. Updated information on cutaneous infection and the definition of a carrier. Updated vaccination information

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