

West Nile Virus

Signs and Symptoms	<ul style="list-style-type: none"> • Most infections asymptomatic (~80%) • Neuroinvasive: aseptic meningitis, encephalitis, acute flaccid paralysis, optic neuritis, and cranial nerve abnormalities (<1%) • Non-neuroinvasive: acute febrile illness; may be headache, muscle or joint aches, rash, or gastrointestinal symptoms (~20%) 	
Incubation	Typically 2-6 days (range 2-14 days); several weeks in immunocompromised individuals	
Case classification (not used for clinical diagnosis)	<p>Clinical criteria:</p> <p>Neuroinvasive: meningitis, encephalitis, acute flaccid paralysis, or other acute signs of central or peripheral neurologic dysfunction</p> <p>Non-neuroinvasive: Fever or chills and absence of neuroinvasive disease</p>	
	<table border="1"> <tr> <td> <p>Confirmed: Clinically compatible with: isolation of virus, WNV antigen or nucleic acid in body fluid, OR four-fold or greater change in serum antibody titers, OR serum IgM confirmed by PRNT. Neuroinvasive only: Above criteria OR clinically compatible with IgM in CSF with a negative result for other arbovirus IgM in CSF</p> </td> <td> <p>Probable: Clinically compatible with IgM in serum but with no other testing</p> <p>Neuroinvasive only: Above criteria OR IgM in CSF but with no other testing</p> </td> </tr> </table>	<p>Confirmed: Clinically compatible with: isolation of virus, WNV antigen or nucleic acid in body fluid, OR four-fold or greater change in serum antibody titers, OR serum IgM confirmed by PRNT. Neuroinvasive only: Above criteria OR clinically compatible with IgM in CSF with a negative result for other arbovirus IgM in CSF</p>
<p>Confirmed: Clinically compatible with: isolation of virus, WNV antigen or nucleic acid in body fluid, OR four-fold or greater change in serum antibody titers, OR serum IgM confirmed by PRNT. Neuroinvasive only: Above criteria OR clinically compatible with IgM in CSF with a negative result for other arbovirus IgM in CSF</p>	<p>Probable: Clinically compatible with IgM in serum but with no other testing</p> <p>Neuroinvasive only: Above criteria OR IgM in CSF but with no other testing</p>	
Differential diagnosis	Different agents difficult to distinguish due to cross-reactivity and persisting IgM (years). Other illnesses: hepatitis, influenza, leptospirosis, malaria, viral rash illness, viral hemorrhagic fever, viral meningitis, cholecystitis	
Treatment	Supportive	
Duration	Generally 3-6 days; may persist for weeks	
Exposure	<p>Vector: mosquitoes, mainly <i>Culex</i> sp. in Washington</p> <p>Reservoirs: birds; humans, horses, and other animals are dead-end hosts</p>	
Laboratory testing	<p>Local health jurisdiction (LHJ) and Communicable Disease Epidemiology (CDE) can arrange testing for borderline commercial result; cases that are neuroinvasive, fatal, or with unusual symptoms; cases with travel to areas with other flaviviruses; other special cases (e.g., pregnant, blood or tissue donor, immunocompromised)</p> <ul style="list-style-type: none"> • Best specimens: generally serum (paired) or CSF • Specimen Collection and Submission instructions https://doh.wa.gov/Portals/1/Documents/5240/SCSI-WNV-MIA-V2.pdf <p><i>Specimen shipping (Section 4):</i></p> <ul style="list-style-type: none"> • Keep specimens cold or if already frozen keep frozen (dry ice), ship according to PHL requirements: https://doh.wa.gov/public-health-provider-resources/public-health-laboratories/lab-test-menu 	
Public health actions	<p>LHJ can consult with CDE 877-539-4344 for testing</p> <ul style="list-style-type: none"> • Confirm diagnosis – confirmatory testing may be needed • Identify potential exposures, particularly local or in the United States • Notify CDE promptly for locally acquired cases (e.g., no out-of-state travel) • Advise no blood donation for 6 months <p><i>Infection Control:</i> standard precautions</p>	

West Nile Virus Disease

1. DISEASE REPORTING

A. Purpose of Reporting and Surveillance

1. To identify areas in which West Nile virus (WNV) is being transmitted.
2. To target public education about reducing mosquito habitats and avoiding mosquito bites.
3. To provide information for mosquito control and environmental health initiatives.
4. To identify periods of time when WNV poses a significant risk to the blood supply.
5. To identify new routes of exposure.

B. Legal Reporting Requirements

1. Health care providers and Health care facilities: notifiable to local health jurisdiction within 3 business days.
2. Laboratories: any positive result excluding IgG notifiable to **local health jurisdiction** within 2 business days; submission on request – specimen associated with any positive result excluding IgG, within 2 business days.
3. Veterinarians: animal cases notifiable to Washington State Department of Agriculture. See: <https://app.leg.wa.gov/WAC/default.aspx?cite=16-70>.
4. Local health jurisdictions: notifiable to Washington State Department of Health (DOH) Communicable Disease Epidemiology (CDE) within 7 days of case investigation completion or summary information required within 21 days.

C. Local Health Jurisdiction Investigation Responsibilities

1. Through investigation, identify unusual exposures and transmission routes other than vector-borne (i.e., donor or recipient of blood products, tissue or organs) within 24 hours of the initial report.
2. Facilitate transport of specimens (e.g., serum or cerebrospinal fluid [CSF]) to the Washington State Public Health Laboratories (PHL) if initial testing or confirmatory testing is needed. Please call CDE prior to submitting specimens (206-418-5500).
3. Report all *confirmed* and *probable* cases and *presumptive viremic donors* to CDE (see definitions below). Complete the West Nile virus case report form <https://www.doh.wa.gov/Portals/1/Documents/5100/210-054-ReportForm-WNV.pdf> and enter the data into the Washington Disease Reporting System (WDRS) as “West Nile Virus Disease” (*Note: do not enter as “Arboviral Disease”*).

2. THE DISEASE AND ITS EPIDEMIOLOGY

Background

West Nile virus (WNV) was first isolated in 1937 from a febrile woman in the West Nile District of Uganda. WNV was first recognized as a cause of severe meningoencephalitis in elderly patients during an outbreak in Israel in 1957. It was first introduced to North America

in 1999 in New York City. Since then, WNV has spread across the United States, and was first known to have reached Washington in 2002.

A. Etiologic Agent

West Nile virus is a single-stranded RNA virus of the family Flaviviridae, genus Flavivirus. It is a member of the Japanese encephalitis virus serocomplex, which contains several medically important viruses that cause encephalitis in humans: Japanese encephalitis, St. Louis encephalitis, Murray Valley encephalitis, and Kunjin. The close antigenic relationship of flaviviruses, particularly those in the Japanese encephalitis complex, accounts for the serologic cross-reactions observed in the diagnostic laboratory.

B. Description of Illness

Serosurveys have shown that less than 1% of WNV-infected persons develop a more severe form of disease, called **WNV neuroinvasive disease (WNND)** which includes encephalitis, meningitis, acute flaccid paralysis (“WNV poliomyelitis”), optic neuritis, and cranial nerve abnormalities. Approximately 20% of infected individuals develop **West Nile fever (WNF)** and 70- 80% remain asymptomatic.

The incidence and case-fatality rate of WNND increase with age, with the greatest risk occurring in persons over 60 years old and some immunocompromised patients (for example, organ transplant patients). Among those with WNND, the overall case-fatality rate is 10%, but is significantly higher for patients with WNV encephalitis and poliomyelitis than WNV meningitis. WNF is a febrile illness, often accompanied by headache, myalgias, fatigue, and malaise. Gastrointestinal symptoms, rash, and lymphadenopathy occasionally occur. Symptoms generally last 3 to 6 days but may persist for weeks. WNF rarely results in death.

C. West Nile Virus Infections in Washington State

Washington State agencies conduct surveillance for WNV infections in humans, birds, mosquitoes, horses and other animals. The first detections of the virus in Washington was in 2002; the first locally acquired human infections were reported in 2006. Since then, 0 to 13 cases per year have typically been reported in Washington, but numbers were higher in 2009 and 2015. During 2009 there were 38 cases and 2 presumptive viremic blood donors reported, almost all exposed in eastern Washington. In 2015, 24 cases were reported, 22 of which were acquired in eastern Washington. Thirteen cases were reported in 2017, and 3 cases were reported in 2018, including the first human case with exposure in western Washington. Visit West Nile Virus Data for current or historical information <https://doh.wa.gov/data-and-statistical-reports/diseases-and-chronic-conditions/west-nile-virus>.

D. Vectors and Reservoirs

WNV is maintained in an enzootic cycle involving vector mosquitoes and many bird reservoir species. Although corvids (crows, ravens, magpies, jays) infected with WNV often become ill and die, many infected birds survive and are asymptomatic. In the United States infections have been found in more than 300 native and exotic bird species. Competent bird reservoirs may have WNV circulating in their bloodstream for 1-4 days after infection. Mosquitoes that feed on them during that period can then be infected.

WNV is transmitted mainly by mosquitoes in the *Culex* subgenus, which occur in

Washington State, though many other mosquito species are also known to become infected. Humans, horses, and most other mammals do not develop the high-level viremia that infect mosquitoes, and are considered "dead-end" or incidental hosts.

E. Modes of Transmission

The main route of transmission is through the bite of an infected mosquito. In very rare cases, WNV has been transmitted through blood transfusions, organ transplants, percutaneous injuries in the laboratory, and from mother to baby via the placenta and possibly breast milk. WNV is not spread through casual contact (e.g., kissing, touching).

F. Incubation Period

Usually 2 to 6 days (but can range up to 14 days).

G. Period of Communicability

Infected people may develop a short lived (2–3 day) low-level viremia that can be found in donated blood; blood collection centers screen donated units to prevent contaminated blood from being used in transfusions.

H. Treatment

In the absence of an effective antiviral agent, treatment for WNV infection is supportive. Treatment for WNNND often involves hospitalization, intravenous fluids, respiratory support, and prevention of secondary infections.

3. CASE DEFINITION

A. Clinical Description

Most West Nile virus (WNV) infections are asymptomatic. Clinical disease ranges from mild febrile illness to severe encephalitis. For the purposes of surveillance and reporting, based on clinical presentation, WNV disease cases are categorized into two primary groups: neuroinvasive disease and non-neuroinvasive disease.

Neuroinvasive disease: WNV can cause neuroinvasive disease such as aseptic meningitis, encephalitis, or acute flaccid paralysis (AFP). These illnesses are usually characterized by the acute onset of fever with headache, myalgia, stiff neck, altered mental status, seizures, limb weakness, or cerebrospinal fluid (CSF) pleocytosis. AFP may result from anterior ("polio") myelitis, peripheral neuritis, or post-infectious peripheral demyelinating neuropathy (i.e., Guillain-Barré syndrome). Less common neurological manifestations, such as cranial nerve palsies, also occur.

Non-neuroinvasive disease: WNV is capable of causing an acute systemic febrile illness that may include headache, myalgias, arthralgias, rash, or gastrointestinal symptoms. Other physical complaints may include vertigo, stiff neck, or muscle weakness without progression to more clinically apparent neurological involvement.

B. Clinical Criteria for Diagnosis

A clinically compatible case of WNV disease is defined as follows:

Neuroinvasive disease

- Meningitis, encephalitis, acute flaccid paralysis, or other acute signs of central or peripheral neurologic dysfunction, as documented by a physician, **AND**
- Absence of a more likely clinical explanation.

Non-neuroinvasive disease

- Fever or chills as reported by the patient or a health-care provider, **AND**
- Absence of neuroinvasive disease, **AND**
- Absence of a more likely clinical explanation.

C. Laboratory Criteria for Diagnosis

Neuroinvasive disease

Confirmatory:

- Isolation of virus from or demonstration of WNV antigen or nucleic acid in tissue, blood, CSF, or other body fluid; **OR**
- Four-fold or greater change in WNV-specific quantitative antibody titers in paired sera; **OR**
- WNV-specific immunoglobulin M (IgM) antibodies in serum with confirmatory* WNV-specific neutralizing antibodies in the same or a later specimen; **OR**
- WNV-specific IgM antibodies in CSF and a negative result for other arbovirus IgM antibodies in CSF for arboviruses endemic to the region where exposure occurred.

Presumptive:

- WNV-specific IgM antibodies in CSF or serum but with no other testing.

Non-neuroinvasive disease

Confirmatory:

- Isolation of virus from or demonstration of WNV antigen or nucleic acid in tissue, blood, or other body fluid, **excluding CSF**; **OR**
- Four-fold or greater change in WNV-specific quantitative antibody titers in paired sera; **OR**
- WNV-specific immunoglobulin M (IgM) antibodies in serum with confirmatory* WNV-specific neutralizing antibodies in the same or a later specimen.

Presumptive:

- Virus-specific IgM antibodies in serum but with no other testing.

***Arboviral serologic assays:** Assays for the detection of IgM and IgG antibodies commonly include enzyme-linked immunosorbent assay (ELISA), microsphere immunoassay (MIA), or immunofluorescence assay (IFA). These assays provide a presumptive diagnosis and should have confirmatory testing performed. Confirmatory testing involves the detection of arboviral-specific neutralizing antibodies utilizing assays such as plaque reduction neutralization test (PRNT).

Note: See “Specimen Collection” section for appropriate timing of specimen collection.

Comments: In some instances, arboviruses from the same genus produce cross-reactive antibodies. In geographic areas where two or more closely related arboviruses occur, it may be epidemiologically important to attempt to pinpoint the infecting virus by conducting cross-neutralization tests using an appropriate battery of closely related viruses. This is essential, for example, in determining that antibodies detected against St. Louis encephalitis virus are not the result of an infection with West Nile virus, or vice versa, in areas where both of these viruses occur (such as eastern Washington State).

West Nile virus IgM antibodies have been documented to persist for up to 500 days in serum. Therefore the presence of IgM antibodies in serum may signify a past infection. Finding WNV IgM antibodies in CSF or a fourfold or greater change in WNV antibody titers between acute- and convalescent-phase serum specimens provides additional laboratory evidence that WNV was the cause of the patient's recent illness. Likewise, IgG and neutralizing antibodies can persist for many years following a symptomatic or asymptomatic infection. Therefore, the presence of these antibodies alone is only evidence of previous infection and clinically compatible cases with the presence of IgG, but not IgM, should be evaluated for other etiologic agents. Clinical and epidemiologic history also should be carefully considered. See Section 4A for additional details.

D. Case Classification (2015)

Confirmed:

- **Neuroinvasive:** a case that meets the clinical criteria and one or more of the confirmatory laboratory criteria for neuroinvasive disease.
- **Non-neuroinvasive:** a case that meets the clinical criteria and one or more of the confirmatory laboratory criteria for non-neuroinvasive disease.

Probable:

- **Neuroinvasive:** a case that meets the clinical criteria and the presumptive laboratory criterion for neuroinvasive disease.
- **Non-neuroinvasive:** a case that meets the clinical criteria and the presumptive laboratory criterion for non-neuroinvasive disease.

Asymptomatic presumptive viremic blood donors (PVDs): PVDs should be entered into WDRS and classified as "Suspect." If WNV illness develops after the PVD is first reported, reclassify the report as a case. See Section 6.

4. DIAGNOSIS AND LABORATORY SERVICES

A. Laboratory Diagnosis

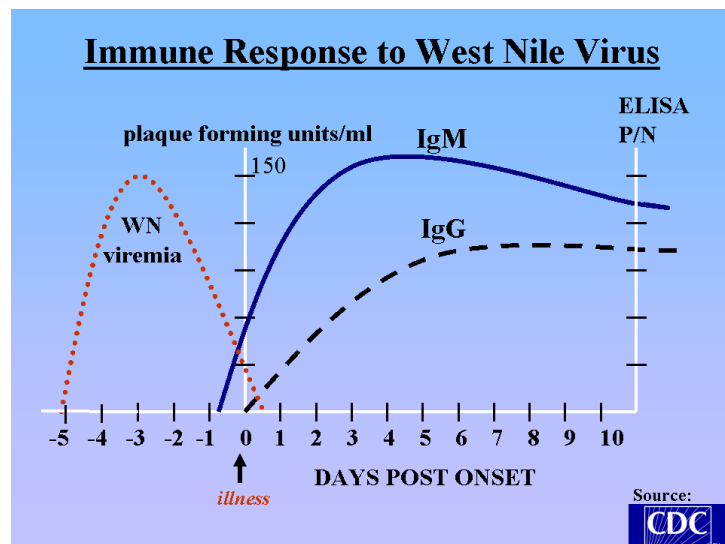
The most efficient diagnostic method for West Nile virus (WNV) infection is detection of virus using the IgM antibody-capture enzyme immunoassay (MAC-EIA) or microsphere immunoassay (MIA) of IgM antibody to WNV in serum collected 8 to 14 days after onset *or* CSF collected within 8 days of illness onset. More than 90% of those infected have detectable serum IgM 8 days after onset; serum collected within 8 days of illness onset may not have detectable IgM and testing should be repeated on a convalescent-phase sample. The EIA can exhibit serologic cross-reactivity in patients who have been recently vaccinated against or recently infected with related flaviviruses (e.g., Japanese encephalitis, yellow fever, or dengue). In addition, since most WNV infections are asymptomatic and IgM can persist in the serum for up to 500 days, the presence of IgM in residents from an endemic area may indicate a previous rather than current infection.

The diagnosis can also be confirmed by a four-fold rise in antibody titer between acute and convalescent (14-21 days after acute) sera. Since serum IgM does not cross the blood-brain barrier, IgM in the CSF strongly suggests central nervous system infection.

The plaque-reduction neutralization test (PRNT) is a more specific test for arthropod-borne flaviviruses and can be used to help distinguish false-positive results from the MAC-EIA or EIA test; however, flaviviruses can still cross-react on PRNT.

Reverse transcription polymerase chain reaction (RT-PCR) assay to detect WNV nucleic acid in serum or CSF is useful for evaluation of patients with immune dysfunction, or to differentiate flaviviruses.

Other antibody detecting diagnostic assays, such as hemagglutination inhibition and indirect fluorescent antibody tests, may be available at commercial laboratories.



B. Services Available at the Washington State Public Health Laboratories (PHL)

Microsphere immunoassay (MIA) for IgM antibody in serum or CSF is currently the only assay available at PHL. This assay is more rapid and specific than the enzyme immunoassay (EIA) test that is used by most commercial laboratories.

Note that PHL require all clinical specimens have two patient identifiers, a name **and** a second identifier (e.g., date of birth) both on the specimen label and on the submission form. Due to laboratory accreditation standards, specimens will be rejected for testing if not properly identified. Also include specimen source and collection date. For details see: <https://www.doh.wa.gov/Portals/1/Documents/5240/SCSI-WNV-MIA-V2.pdf>

C. Criteria for Testing WNV Specimens at PHL

All specimens need to be pre-approved by Communicable Disease Epidemiology (CDE) prior to submission.

Initial testing should be performed at a commercial laboratory. PHL will retest positive specimens in the following circumstances:

1. Persons with commercial laboratory results that indicate only borderline levels of IgM. For instance, PHL would retest if the commercial reference range is >0.9 and the test

result is 1.5. Note that different tests have different reference values. The exact values should be discussed with CDE on a case-by-case basis in the context of exposure and clinical symptoms.

2. Cases of neuroinvasive disease
3. Fatal cases presumed to be WNV.
4. Cases with unusual symptoms.
5. Cases with travel to areas that have other circulating flaviviruses
6. Special cases such as:
 - Pregnant or breastfeeding women symptomatic with suspected WNV infection and their neonates or breastfeeding infants.
 - Recent blood, tissue, or organ donors or recipients suspected to have WNV infection.
 - Persons with known immunocompromising conditions

In cases where the laboratory evidence is inconclusive at PHL, specimens will be forwarded to CDC. The turn-around time for results from CDC is typically 4-6 weeks.

D. Specimen Collection and Shipping

1. Submit ≥ 2 mL of CSF and/or ≥ 1 mL serum (separated serum, not whole blood or plasma).
 - a. Serum should ideally be obtained ≥ 8 days after onset of symptoms. A second serum specimen will be requested if the first is non-reactive or indeterminate and was obtained less than 8 days after onset of symptoms. Convalescent serum should be drawn 14-21 days after the acute specimen.
 - b. CSF should ideally be collected 3-8 days after onset. CSF obtained less than 3 days after onset of symptoms will be accepted, however, if non-reactive, the test does not rule out WNV infection and a serum specimen obtained 8 days after onset will be requested.

Specimens should be refrigerated and transported cold (on regular ice packs). Avoid repeated freeze-thaw cycles. If the specimen is already frozen, ship it frozen (on dry ice). Specimens should be submitted according to PHL requirements:

<https://doh.wa.gov/public-health-provider-resources/public-health-laboratories/lab-test-menu>

5. ROUTINE CASE INVESTIGATIONS

Interview the case or others who may be able to provide pertinent information.

A. Evaluate the Diagnosis

Review the laboratory report, clinical description, and epidemiologic factors such as season and known WNV activity in the area. If commercial laboratory testing is IgM positive, Communicable Disease Epidemiology (CDE) may request facilitating transport of that or another specimen to PHL for further testing.

B. Case Management

1. Hospitalized patients should be treated with standard precautions.
2. Cases do not require isolation.
3. Infected persons should be advised not to donate blood, tissues or organs for at least 6 months following recovery.
4. Infected lactating women should discuss with their medical care provider the risks and benefits of breast-feeding.
5. If the patient received blood products, organs, or tissues in the 30 days prior to onset, contact CDE immediately and inform the blood or tissue bank of the potential source.

C. Identify Potential Sources of Infection

Obtain a complete travel history and history of mosquito bites during the 15 day period prior to symptom onset. Ascertain whether the case received blood products, tissues or organs within 30 days of their WNV infection, and if so, contact CDE immediately and inform the blood or tissue bank of the potential source.

D. Identify Potentially Exposed Persons

Determine if the patient donated blood, tissues or organs, breastfed, or gave birth during the communicable period. If the patient donated blood, tissues or organs in the 30 days prior to onset, contact CDE immediately and inform the blood or tissue bank of the potential blood contamination.

Identify others who may have been exposed at the same time as the patient.

E. Management of Contacts and Other Exposed Persons

No follow up is needed for household and other close contacts since WNV is not transmitted person-to-person, except for rare cases of transplacental transmission, organ transplantation, or blood transfusion. If the patient donated blood or organs, notify CDE and inform the blood or tissue bank of the potential exposure. In cases of potential mother-to-infant transmission, notify CDE and monitor the infant for compatible signs and symptoms for 14 days after last possible exposure. Additional testing may be requested to confirm the transmission.

If the patient donated blood products, organs or tissues in the 30 days prior to onset, contact CDE immediately and inform the blood or tissue bank of the potential exposure.

Instruct others persons with recent mosquito bites in areas where WNV is circulating to seek medical care if they develop fever, headache, rash, or stiff neck.

F. Environmental Evaluation/Management

Notify local environmental health program and/or vector control of locally acquired cases so that they may determine appropriate intervention measures. Environmental measures to reduce WNV transmission may include the elimination of mosquito breeding habitats and the use of chemical (i.e., pesticide) and biological controls. In outbreak settings, an investigation may assist in identifying and controlling factors favoring transmission.

6. MANAGING SPECIAL SITUATIONS

A. Presumptive Viremic Donor (PVD)

1. Blood collection agencies routinely screen blood products for West Nile virus (WNV) using nucleic acid-amplification tests (NAT) during months when there is WNV activity.
2. Blood collection agencies report persons whose blood screens positive to either the local health jurisdiction (LHJ) or to DOH Communicable Disease Epidemiology (CDE). Reports to CDE are forwarded to the LHJ. Local public health professionals should initiate an investigation using the WNV Case Report Form.
<https://www.doh.wa.gov/Portals/1/Documents/5100/210-054-ReportForm-WNV.pdf>
3. Persons whose blood donation screens positive for WNV, with either (a) a single reactive NAT with a signal-to-cutoff (S/CO) value ≥ 17 or (b) two reactive (positive) NATs using an assay which doesn't provide an S/CO value, or (c) two positive with a S/CO value between 1-17, are considered presumptive viremic donors and no further testing is indicated.
4. If a person's blood donation screens positive for WNV using a less stringent method, the LHJ should assist in obtaining a serum sample drawn 8-14 days after the date of the donation to test for IgM. If IgM is detected and the person remains asymptomatic, the person is a PVD. If IgM is not detected, the NAT should be considered a false positive and the investigation should be closed without need for reporting.

Note: Clinical Syndrome and Case Classification: All PVDs should be entered into WDRS, classified as "Suspect," and reported to CDE unless WNV illness is documented. If WNV illness develops after the PVD is first reported, please revisit WDRS to reclassify the patient as either a probable or confirmed case (see Section 3D).

7. ROUTINE PREVENTION

A. Immunization Recommendations

Currently there is no human West Nile virus vaccine available.

B. Prevention Recommendations

1. Reduce exposure to mosquitoes.
 - Make sure windows and doors are "bug tight." Repair or replace screens.
 - Stay indoors at dawn and dusk, if possible, when mosquitoes are most active.
 - Wear a long sleeve shirt, long pants, and a hat when going into mosquito-infested areas, such as wetlands or woods.
 - Use mosquito repellent when necessary. The most effective mosquito repellents contain the EPA approved active ingredients DEET (N, N-diethyl-m-toluamide), Picaridin, oil of lemon eucalyptus, or IR3535. Read and follow label instructions. Permethrin is another long-lasting repellent that is intended for application to clothing and gear, but not directly to skin. In general, the more active ingredient (higher concentration) a repellent contains, the longer time it protects against mosquito bites. Do not over use repellents. Take special care when using repellents on children.

- Additional information regarding the use of mosquito repellents can be found on the CDC website at: <https://www.cdc.gov/mosquitoes/prevention/index.html>.
2. Reduce the number of mosquitoes in breeding sites outdoors by draining sources of standing water.
 - Empty anything that holds standing water (e.g., tires, buckets, plastic covers, toys).
 - Change water in your birdbaths, fountains, wading pools, pet bowls, and animal troughs at least twice week.
 - Recycle unused containers that may collect water—bottles, cans, and buckets.
 - Make sure roof gutters drain properly and clean clogged gutters in the spring and fall.
 - Fix leaky outdoor faucets and sprinklers.

C. Prevention of Laboratory-Associated Infections

Laboratory-associated infections have been reported, attributed to both aerosol and parenteral inoculations. Biosafety Level 2 practices are recommended for diagnostic specimens. Biosafety Level 3 practices are needed for WNV cultures and PRNT.

ACKNOWLEDGEMENTS

This document is a revision of the Washington State Guidelines for Notifiable Condition Reporting and Surveillance published in 2002 which were originally based on the Control of Communicable Diseases Manual (CCDM), 17th Edition; James Chin, Ed. APHA 2000. We would like to acknowledge the Oregon Department of Human Services for developing the format and select content of this document.

UPDATES

March 2008: In Section 1C, the guideline for timeliness of initiating an investigation was removed.

July 2008: In Section 1C, the CDC WNV Enhanced surveillance form was added. In Section 4A, additional information was added regarding laboratory tests. In Section 8B, IR3535 was added as an EPA approved effective mosquito repellent.

June 2009: In Section 2B, case-fatality rates and new terminology for clinical syndromes were updated. In Section 3C, case classification for presumptive viremic donors was updated. In Section 4B, the change in laboratory tests available at PHL were noted. In section 4D, information about the timing of specimen collection was clarified. In Section 8C, prevention of lab-associated infections was moved from Section 4.

August 2009: In section 1C, request to complete a separate CDC enhanced surveillance form was removed.

May 2010: Case definitions (Section 3) and criteria for testing at PHL (Section 4C) were updated.

January 2011: The Legal Reporting Requirements section has been revised to reflect the 2011 Notifiable Conditions Rule revision. Section 3 was nearly completely replaced to reflect the 2011 CSTE case definition and required laboratory evidence changes. Changes to the type of testing available at WA PHL were made in Section 4B.

September 2014: Section 3 was revised to reflect the 2014 CSTE case definition, in which the requirement for documented temperature of fever was changed to allow subjective fever or chills. For arboviral diseases (other than Dengue), fever has been removed entirely from the criteria for neuroinvasive disease; it no longer needs to be measured for non-neuroinvasive disease. The laboratory criteria were differentiated for neuroinvasive disease and non-neuroinvasive disease. Previous sections 5 and 6 were combined into a single section 5.

March 2017: Front page added, Sections 2B and C and 4C were updated to reflect current knowledge and recommendations.

Dec 2019: Links and Sections 2 B and C updated.

December 2022: Section 1B.3 lab reporting guidelines updated by WAC 246 101 201 Jan 2023 standards.

December 2023: For 2024 WAC revision updated laboratory submission.

June 2024: CDC links updated.

To request this document in another format, call 1-800-525-0127. Deaf or hard of hearing customers, please call 711 (Washington Relay) or email doh.information@doh.wa.gov.